Anti-Browning of Mushroom (*Agaricus bisporus*) Slices by Glutathione during Hot Air Drying

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Abstract: Browning of mushroom tends to occur during hot air drying due to Poly Phenol Oxidase (PPO), while glutathione is known for its ability to inhibit the activity of PPO and browning. In this study, the efficacy of glutathione in inhibiting browning on mushroom slices was estimated. Browning of mushroom slices treated with glutathione was monitored during hot air drying. PPO activity in mushroom was inhibited by 98.2 with 0.08% glutathione. Compared with the control, mushroom slices treated with glutathione showed no browning during hot air drying. These results indicate that application of glutathione is a promising method of Anti-browning of mushroom by glutathione during hot air drying.

Keywords: Browning, drying, glutathione, mushroom

INTRODUCTION

Mushrooms, a special group of macroscopic fungi, have been used as a food source since ancient times. Mushrooms are liked for their delicious flavor, low calorific value, high protein contents (20-40%), vitamins of B-group and minerals (Walde et al., 2006).

Mushrooms are highly perishable commodities and they start deteriorating immediately within a day after harvest. In view of their highly perishable nature, the fresh mushrooms have to be processed to extend their shelf life for off-season use. Among the various methods employed for preservation, drying is one of the common methods used for mushrooms (Argyropoulos et al., 2011; Kar et al., 2004; Kotwalwale et al., 2007; Kumar et al., 2013; Walde et al., 2006).

Colour of the product is of prime importance to the consumer as product quality criteria (Matser et al., 2000). Thus, browning in mushrooms during drying, which is ascribed to the activity of phenolic compounds, oxygen and Poly Phenol Oxidase (PPO), decreased the sensory quality and commercial value of the dried mushrooms (Kotwalwale et al., 2007; Kumar et al., 2013). Though application of advanced drying methods, e.g., microwave drying and freeze-drying, can alleviate this problem, it needs expensive equipments and high production cost. Therefore, using color protecting agents is a practical way to inhibit browning in mushrooms during drying.

Glutathione is known for its ability to inhibit the activity of PPO and browning (Jiang and Fu, 1998). Thus, the aim of this study is to investigate the inhibitory effects of glutathione on the activity of PPO and browning in mushroom during hot air drying.

MATERIALS AND METHODS

Materials: Fresh mushrooms (*Agaricus bisporus*) were purchased from a local supermarket and stored at 4°C until used. Glutathione was purchased from Fuchen Chemical reagents Co. (Tianjin, China). L-β- (3, 4-dihydroxyphenyl) alanine (L-DOPA), Brij-35, sodium phosphate and ammonium sulphate were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Preparation of PPO extract from mushroom: Mushroom (100 g) was powdered by grinding with liquid nitrogen in a Waring blender (JJ-2, Wuxi Woshin Instruments Co., Ltd. and Wuxi, China). The powder was kept in a polyethelene bag and stored at -20°C until use (not more than 24 h). The isolation of PPO was carried out according to the method described by Nirmal and Benjakul (2009a) with slight modifications. The powder (50 g) was mixed with 150 mL of the extracting buffer (0.05 M sodium phosphate buffer, pH 7.2, containing 1.0 M NaCl and 0.2% Brij-35). The mixture was stirred continuously at 4°C for 30 min, followed by centrifugation at 8000×g and 4°C for 30 min using a refrigerated centrifuge (GTR10-1, Beijing Era Beili Centrifuge Co., Ltd., Beijing, China). Solid ammonium sulphate was added into the supernatant to obtain 40% saturation and allowed to stand at 4°C for 30 min. The precipitate was collected by centrifugation at 12,500×g at 4°C for 30 min using a refrigerated centrifuge. The pellet obtained was dissolved in a minimum volume of 0.05 M sodium phosphate buffer, pH 7.2 and dialyzed against 15 volume of the same buffer at 4°C with three changes of dialysis buffer. The insoluble materials were removed by centrifugation at 3000×g at 4°C for 30 min and the supernatant was used as “crude PPO extract”.

Preparation of mushroom slices: Fresh mushrooms were cut into slices of uniform size (4×4 mm). The slices were divided into four groups, namely, control, 0.01%, 0.03% and 0.06% glutathione treatment groups. Glutathione was dissolved in deionized water and filtered through Whatman filter paper (pore size, 0.45 mm). The sliced mushrooms were immersed in different concentrations of glutathione solution for 1 h at 4°C, followed by washing with deionized water.

Browning of mushroom slices: The slices were prepared as follows: 1 g of mushroom slices were mixed with 1 mL of deionized water and 0.5% Brij-35 at 4°C. The mixture was filtered through Whatman filter paper (pore size, 0.45 mm). The filtrate was used as the source of PPO. The PPO activity was measured using the method described by Nirmal and Benjakul (2009a). The absorbance of the reaction mixture was measured at 420 nm using a spectrophotometer (UV-1601, Shimadzu, Japan) and the PPO activity was calculated as nmol of DOPA oxidized per min per g of mushroom.

Browning of mushroom slices was estimated using a similar method to that described by Nirmal and Benjakul (2009a). A 6% dextrose solution and 3% lactic acid were used as the browning agent. The dextrose and lactic acid solutions were adjusted to pH 3.5 with 2 M sodium hydroxide and 3 M hydrochloric acid, respectively. The mushroom slices were immersed in 10 mL of dextrose and lactic acid solutions for 24 h at 25°C. The absorbance of the reaction mixture was measured at 420 nm using a spectrophotometer (UV-1601, Shimadzu, Japan) and the browning index was calculated as nmol of DOPA oxidized per min per g of mushroom.
Determination of PPO activity: The activity of PPO was determined using L-DOPA as a substrate according to the method described by Nirmal and Benjakul (2009b) with a slight modification. The assay system consisted of 100μL of crude PPO extract, 600 μL of 15 mM L-DOPA in deionised water, 400 μL of 0.05 M phosphate buffer (pH 6.0) and 100 μL of deionised water. The PPO activity was determined for 3 min at 45°C by monitoring the formation of dopachrome at 475 nm using a UV spectrophotometer (752, Nanjing Qilin Instruments Co., Ltd., Nanjing, China). One unit of PPO activity was defined as an increase in the absorbance by 0.001 at 475 nm/min/mL. Enzyme and substrate blanks were prepared by excluding the substrate and enzyme, respectively, from the reaction mixture and deionised water was used instead.

Effect of glutathione on the activity of PPO: Glutathione was dissolved in distilled water to various concentrations (0.04, 0.08, 0.12, 0.16, 0.20 and 0.24%, respectively, w/v). Glutathione solution (100 μL) was mixed with crude PPO extract (100 μL) and the mixture was incubated at room temperature for 30 min and then, the assay buffer (400 μL, 0.05 M phosphate buffer, pH 6.0) was added. To initiate the reaction, 600 μL of preincubated 15 mM L-DOPA (45°C) were added. The reaction was conducted at 45°C and the absorbance at 475 nm was monitored for 3 min. The control was run in the same manner, except the deionized water was used instead of glutathione solution. The sample blank was prepared by using distilled water instead of L-DOPA. One unit of PPO activity was defined as that causing an increase in the absorbance by 0.001 at 475 nm/min/mL. Residual activity was determined and expressed as the activity relative to the control (without glutathione) as follows:

Relative activity (%) = (B/A) × 100

where,
A: PPO activity of control
B: PPO activity in the presence of glutathione (Nirmal and Benjakul, 2009a)

Treatments of mushroom: Fresh mushrooms with uniform size and color were washed with tap water, destemmed and cut into slices with equal thickness using a sharp stainless steel knife. All slice cut from each mushroom was pooled together and treated in one of the dipping solutions (water for control and 0.08% glutathione solution) for 5 min, drained on a clean paper towel, taken for drying and spread over perforated aluminum trays and trays were kept in the drying chamber at different temperatures (50, 55, 60, 65, 70 and 75°C, respectively) for varying time (20, 40, 60, 80, 100, 120, 140, 160 and 180 min, respectively). During the drying, color and moisture content of the sample was recorded.

Color and moisture determination: The color of mushroom slices was measured using a Minolta colorimeter (CS-100A, Minolta Co. Ltd., Japan) according to Lu et al. (2007) with slight modifications (2007). The degree of browning was expressed by the changes in the Lightness (L) value. Moisture content of mushroom was determined by vacuum oven method.

Statistical analysis: All of the tests were performed in triplicate and data were reported as means±S.D. Analysis of variance and significant differences among means were tested by one-way ANOVA using SPSS software (version 17.0 for Windows, SPSS Inc., Chicago, IL, USA).

RESULTS AND DISCUSSION

Effect of glutathione on PPO inhibition: Glutathione is the most abundant non-protein thiol compound present in living organisms and used as a pharmaceutical compound and can be used in food additives and the cosmetic industries (Li et al., 2004). It was used for the inhibition of PPO in apple (Billaud et al., 2004; Brun-Mérimée et al., 2004; Gacche et al., 2004), Loquat Fruit (Ding et al., 1998), litchi fruit (Jiang and Fu, 1998), must (Cheynier et al., 1990) and

Fig. 1: Effect of glutathione on inhibition of PPO activity of mushroom
Fig. 2: Effect of glutathione on inhibition of browning formation and moisture content in mushroom during hot air drying at 70°C for varying time

Fig. 3: Effect of glutathione on inhibition of browning formation in mushroom during hot air drying at different temperatures for 120 min

Fruit juices and protein-containing foods (Molnar-Perl and Friedman, 1990).

The effect of glutathione on the inhibition of PPO in mushroom is shown in Fig. 1. The relative activity of PPO decreased sharply with the increase in the level of glutathione up to 0.06% in the reaction mixture and decreased slowly with the amount of glutathione between 0.06 and 0.08%. No decrease in relative activity of PPO was observed when the amount of glutathione was further increased, indicating that at 0.08% of glutathione, all PPO was saturated by the glutathione. It is therefore that the optimum amount of glutathione under this condition was 0.08% (Fig. 1). Therefore, 0.08% (w/v) glutathione, which shows the highest PPO inhibitory effect, was used for further study.

Effect of glutathione on inhibition of browning formation in mushroom during hot air drying:
Browning formation decreases the sensory quality of mushroom products. Recently, many antioxidants have been applied to the anti-browning of mushroom, such as ascorbic acid (Zhang et al., 2004), ascorbic acid derivatives (Hsu et al., 2006), cysteine (Dorantes-Alvarez et al., 1998; Huang et al., 2008), tetra sodium pyrophosphate (Dorantes-Alvarez et al., 1998), Benzoic Acid, Glutathione, EDTA, 4-Hexylresorcinol and Sodium Chloride (Weemaes et al., 1999). However, using glutathione to inhibit browning formation of mushroom during hot air drying is not frequently reported. Effect of glutathione on the anti-browning of mushroom during hot air drying at 70°C for varying time is shown in Fig. 2. Both control and 0.08% glutathione samples showed identical drying characteristics: the moisture content in both samples decreased to the lowest levels in 120 min; there was no significant difference in the final moisture contents between the two samples (~9%, p>0.05). The L values of control and 0.08% glutathione samples were 74 and 51 on 120 min at 70°C, respectively (Fig. 2). The L values of control samples decreased with the decreasing of the moisture contents within 120 min and decreased further after 120 min even the moisture maintained unchanged. However, the L values of 0.08% glutathione samples did not decrease significantly during the whole drying time course (p>0.05).

Effect of glutathione on the anti-browning of mushroom during hot air drying at different temperatures for 120 min is shown in Fig. 3. The L
values of control samples decreased with the increasing of temperature. In the case of 0.08% glutathione samples, the $L$ values did not affected significantly by drying temperature ($p>0.05$). The results indicate that higher drying temperature could be used to dry mushroom pretreated with 0.08% glutathione to get higher drying efficiency. The color quality of control and 0.08% glutathione samples is shown in Fig. 4. For 0.08% glutathione samples, color showed faint yellow features (Fig. 4a), while that of the control got serious gray and brown (Fig. 4b). Results showed that glutathione as anti-browning agent prevented mushroom slices from browning during hot air drying, indicating that glutathione is an effective anti-browning agent. Previous studies indicated that glutathione inhibits browning in apple (Jiang and Fu, 1998). The inhibition of glutathione of mushroom browning coincided with the ability of the treatment to inhibit PPO activity (Fig. 1).

CONCLUSION

Glutathione treatment could be used to inhibit PPO activity in mushroom and the efficacy was associated with dose. PPO activity in mushroom was inhibited by 98.2 with 0.08% glutathione. Pretreatment with 0.08% glutathione could be used to prevent browning from forming in mushroom during hot air drying, while that of the control got serious gray and brown. Therefore, pretreatment with 0.08% glutathione means high quality of dehydrated mushrooms.

REFERENCES


